

# Standard Operating Procedure

## CAGE TRAPS FOR LIVE CAPTURE OF TERRESTRIAL VERTEBRATES

**Animal welfare is the responsibility of all personnel involved in the care and use of animals for scientific purposes.**

**Personnel involved in an Animal Ethics Committee approved project should read and understand their obligations under the *Australian code for the care and use of animals for scientific purposes*.**

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# 1 Purpose

Cage trapping is a common method used for monitoring many species of small to medium-sized mammals. Cage traps usually operate using a treadle plate mechanism, which is set off when an animal steps on the elevated trigger plate and springs the door closed. Cage traps will also catch a range of non-target species including birds and reptiles.

This Standard Operating Procedure (SOP) provides advice on the use of cage traps for non-lethal trapping of terrestrial vertebrate fauna.

# 2 Scope

This SOP has been written specifically for scientific and education purposes, and endorsed by the Department's Animal Ethics Committee. However, this SOP may also be appropriate for other situations.

This SOP applies to all fauna survey and monitoring activities that may require the use of cage traps undertaken across the State by Department of Biodiversity, Conservation and Attractions (hereafter Department) personnel. It may also be used to guide fauna monitoring activities undertaken by Natural Resource Management groups, consultants, researchers and any other individuals or organisations. All Department personnel involved in the use of cage traps should be familiar with the content of this document.

Projects involving wildlife may require a licence under the provisions of the *Wildlife Conservation Act 1950* and/or the *Biodiversity Conservation Act 2016*. Personnel should consult the Department's Wildlife Licensing Section and Animal Ethics Committee Executive Officer for further guidance. In Western Australia any person using animals for scientific purposes must also be covered by a licence issued under the provisions of the *Animal Welfare Act 2002*, which is administered by the Department of Primary Industries and Regional Development. This SOP complements the *Australian code of practice for the care and use of animals for scientific purposes* (The Code). The Code provides governing principles to guide decisions and actions of personnel involved in the care and use of animals, and contains an introduction to the ethical use of animals in wildlife studies. A copy of The Code may be viewed by visiting the National Health and Medical Research Council website (<http://www.nhmrc.gov.au>).

# 3 Animal Welfare Considerations

To reduce the level of impact of cage trapping on the welfare of animals, staff must consider, address and plan for the range of welfare impacts that may be encountered. Strategies to reduce impacts should be identified during the planning stage to ensure that they can be readily implemented during trap set up and trap checking and contingencies for managing welfare issues have been identified. All handlers and volunteers involved in the project should be aware of the range of issues that they may encounter, the options that are available for reducing impacts and improving animal welfare, and the process for managing adverse events.

Department projects involving cage trapping will require approval from the Department's Animal Ethics Committee.

Key animal welfare considerations that should be considered when cage trapping are listed below and highlighted throughout the document.

### 3.1 Injury and unexpected deaths

If adverse events including injury, unexpected deaths or euthanasia occur then it is essential to consider the possible causes and take action to prevent further incidents. For projects approved by the Department's Animal Ethics Committee, adverse events must be reported in writing to the AEC Executive Officer as soon as possible after the event by completing an *Adverse Events* form. Guidance on field euthanasia procedures is described in the Department SOP for *Humane Killing of Animals under Field Conditions*. Where disease may be suspected, refer to the Department SOP for *Managing Disease Risk in Wildlife Management* for further guidance.

### 3.2 Level of impact

Potential animal welfare impacts of cage trapping include:

- Capture myopathy (particularly for Macropods)
- Trauma (e.g. head or nose injuries from hitting walls of the trap)
- Smaller non-target species stuck in mesh
- Stress as a result of harsh environmental conditions within the trap (i.e. temperature).
- Distress (caused by confinement, discomfort, social isolation, separation of mother and young, exposure to predators, ants, etc.)
- Predation

If the cage traps are properly monitored and preventative actions are utilised then the impact should be small and only short-term.

## 4 Approved Trap Types

Large Cage: Galvanised wire mesh cage trap (approx. 45cm x 45cm x 90cm) with a treadle plate release mechanism. Large cages are used primarily for feral cat (*Felis catus*) trapping.

Small Cage: Galvanised wire mesh cage trap (20cm x 20cm x 56cm) with a treadle plate release mechanism. Collapsible forms are available. Used for most medium-sized mammals such as chuditch (*Dasyurus geoffroii*), quenda (*Isoodon obesulus fusciventer*), brushtail possums (*Trichosurus vulpecula*) and woylies (*Bettongia penicillata ogilbyi*). Small cages also catch small Dasyurids and rodents as well as Varanids, large skinks and occasionally birds.

Some old style traps used a trigger mechanism attached to a bait hook hanging from the roof of the trap, which when tugged on, releases the door, or hooks were simply included to keep bait off ground. Traps with hook-release mechanisms are not acceptable. Where traps with bait hooks are still in use, the hook must be completely closed to form a loop so that an animal is unable to get caught on the hook.

Many cage traps used in Western Australia are manufactured by Sheffield Wire Products (Sheffield Rd, Welshpool WA) and so are sometimes referred to as "Sheffields." Cage traps

manufactured with different trigger mechanisms may also be appropriate and their use is not excluded, provided they do not pose additional welfare risks to animals (see reference to 'hooks' above). Projects approved by the Department's Animal Ethics Committee preferring to use alternative cage style traps to those mentioned here may do so if they describe in detail the differences in design and are able to report on the survivorship rates and the welfare impacts.

All traps should be checked for sharp edges, protrusions, or gaps/holes large enough for entrapment of digits/limbs which can cause injury, regardless of age of trap (some new traps can have rough or sharp edges from the milling/cutting process.) Proper function of the doors and trigger mechanisms should also be checked as malfunctioning devices may pose a risk by partially trapping an animal.

The solid nature of cage traps means that animals can injure themselves whilst inside the trap. To minimise these injuries soft trap options have been developed and are covered in the Department SOP for *Soft Cage Traps for Capture of Macropods*. These soft traps are preferred for species that are particularly prone to injury or capture myopathy and have been effectively used for a variety of species including rock-wallabies (*Petrogale lateralis*), tammar wallabies (*Notamacropus eugenii derbianus*) and mala (*Lagorchestes hirsutus*).



Figure 1 A cage trap with hessian and vegetation cover. Photo: Christine Freegard/DBCA

## 5 Procedure Outline

### 5.1 Setting and positioning traps

(a) The location and configuration of trap placement (i.e. transect or grid) as well as the number of traps will be determined by the purpose of the study and should be planned before commencing the survey. Consider the target species' likely use of habitat and home range and welfare implications of trap placement when designing trap configuration and layout.

*Example:* Transects of 50 small cage traps spaced at 200m intervals (total 10km) have been used as the standard method for monitoring target species under the Western Shield program.

(b) Trap locations must be marked so that no traps are missed when checking or removing them (e.g. with flagging tape which is labelled and using a numbering system which uniquely identifies each trap). A GPS reading for each trap point is strongly recommended. Permanent

monitoring trap sites should also be marked using a permanent marker (e.g. numbered dropper post). The location information for permanent monitoring transects and their trap points should be recorded on datasheets and a database.

(c) If setting traps along roads or vehicle tracks, the traps must be set so that they minimise the impact on the animals. Traps should be placed away from the roadside (generally a distance of 5m or greater on publicly accessible roads and tracks) so that they are not readily visible from the road to avoid public curiosity and possible theft of traps, and to reduce the disturbance on trapped animals from passing vehicles.

(d) Trap placement:

**ANIMAL WELFARE:** Trap placement can greatly affect animal welfare. Consider the climate of the area you are trapping in and the species biology (e.g. thermoregulation characteristics) when choosing a trap position. Traps need to be placed in suitable locations that provide shelter from the sun and protection from rain to reduce exposure of trapped animals. For example, consideration needs to be given to the movement of the sun (and therefore shade), prevailing winds and drainage in wet conditions. Consider the orientation of the sun and the period of the day when the captured animal will be in the trap.

Thick trap covers that provide protection from the elements and reduce the sense of exposure by the animals are required (refer point (f) below). If the traps are likely to capture species that are prone to panic or stress (e.g. woylies), trap placement should also allow animal handlers to approach the traps without increasing the stress of the animal (e.g. approach trap from the side rather than the front, reduced noise from walking on leaf litter, and minimal talking).

Do not place traps on or in the vicinity of ant nests.

(e) Traps must be set in level positions using natural cover wherever possible. Debris and/or vegetation should be cleared from under the trap to ensure stability and prevent obstructions from stopping the dropping and locking of the trap door. In some areas where the ground is uneven or ground vegetation makes it difficult for the door to close, it may be appropriate to place a short straight stick, no longer than the width of the cage, under the bottom front edge of the trap to lift the bottom lip of the trap mouth just off the ground and provide clearance for the door to close easily. Note that this must be done in a way that won't reduce trap stability and won't create issues for an animal stepping into the trap.

(f) Cage traps must have adequate shelter and protection for the welfare of captured animals. Cage traps should be covered with heavy weight hessian (or similar material with the same protective qualities) to provide captured animals with security and shelter from the elements. Place the hessian over the top of the trap and wrap around to cover exposed sides. The cover needs to be secured to ensure that it cannot be easily removed by an animal and wind cannot blow the hessian off the trap. Options include using a rock or log, nestling the trap into a bush, tucking the edges of the bag under the trap (ensuring the stability of the trap is not impeded) or piling sand on the edges of hessian. Ensure that the trap release mechanism is not impeded by the hessian or method used to secure it.

(g) Before the trap is left, it is important to re-check that the mechanism is working properly, the trap cover is effective and secure, and the trap is positioned to take advantage of shade in the morning. Faulty equipment reduces the opportunity to trap animals and can result in poor data and reduce the value of the trapping effort.

- (h) All traps must be accounted for before and after each trapping session.

## 5.2 Baiting traps

When choosing the type of bait for your traps always consider the target species and possible non-target captures. Bait is intended to lure an animal into the trap and for some species, provides a small amount of food while the animal is trapped.

**Small Cage:** The standard bait used in small cages is a mixture of peanut paste and rolled oats which is also known as “universal bait” (*Note: sardines may increase the attraction of ants and you may want to consider excluding them from the bait if ants are an issue*). Small cages require a quantity about half to a third of the size of a golf ball. Refer to Appendix II for more information.

Alternative baits such as tuna, sardines, chicken and bacon can be considered when targeting carnivorous mammals such as chuditch. Use of a meat bait may also increase captures of reptiles, particularly varanids and skinks.

If using a meat bait, personnel should maintain good hygiene practises when handling the baits, such as washing and disinfecting their hands after contact with the bait, and avoiding touching their face, mouth and trapping equipment until cleaned. Incorrect handling and hygiene surrounding meat baits can potentially lead to foodborne viruses and infections, such as Salmonella. Gloves can also be worn when preparing or handling meat baits. Gloves should be removed when no longer handling baits to avoid contaminating other equipment.

Other bait types or ingredients may be used if these have been identified as appropriate and approved for use for a particular project and/or species.

Care must be taken when baiting traps to ensure that the bait is placed clear of the treadle plate and does not impede the closing mechanism. To avoid bait rolling underneath the treadle plate and rendering the trap inoperative, it is recommended that universal bait balls are slightly squashed so that they cannot roll under the treadle.

Baits should be replaced when their effectiveness as a lure is reduced (e.g. when the odour of the bait is reduced or gone) or if the bait may impact on an animal’s health if consumed (e.g. rancid). Baits should be replaced rather than additional baits placed in the cage – more bait in the cage may increase the probability of the treadle being impeded.

## 5.3 Checking traps

**ANIMAL WELFARE:** In determining the duration and frequency of trapping you should consider the purpose of your study and the potential welfare impacts from recapturing animals on multiple occasions (e.g. limitations on feeding, welfare of dependent young). Consider the duration and frequency that will allow the goal of the activity to be achieved with the minimal impact on animals. Some animals become “trap happy” (entering traps on multiple consecutive nights) and this can impact their wellbeing by disrupting behaviours such as normal feeding, foraging, breeding and defending territories. This is particularly relevant to small mammals (e.g. honey possums) which due to their small size, are at risk of death if prevented from feeding. Where honey possums are prevalent, a sugar solution (e.g. Spark liquid) should be available when checking traps.

Avoid trapping in breeding seasons where lactating females may be separated from dependent young or when there is an increased likelihood of injury or separation of

dependent young (e.g. brushtail possums during pouch emergence). However, many species breed throughout the year making it impossible to completely avoid trapping animals at sensitive times. If captured, lactating animals should be released as soon as possible. If the same lactating female is caught on successive nights, consideration should be given to moving or closing the trap.

Avoid trapping or close traps in extreme weather conditions. Plan ahead and monitor long-range and daily weather forecasts.

For programs such as *Western Shield* monitoring it is recommended that traps are set for a minimum of four consecutive nights.

(a) All traps must be accounted for during each day's trapping. Personnel undertaking the trapping should keep tallies of traps to ensure that all are checked. This is the responsibility of the person in charge at the survey location on the day. There is no excuse for leaving traps unchecked.

**ANIMAL WELFARE:** The timing and frequency of trap checking and clearing should be determined by considering the behaviour and biology of the target species (and potential by-catch species) in association with the environmental conditions at the site. Trap checking timing and frequency should be reviewed and adapted when and if conditions change or adverse events occur. Traps may need to be checked more frequently throughout the day and/or night if prolonged trap confinement or environmental conditions are likely to increase the impact on animal welfare and affect survivorship.

(b) Where nocturnal species are being targeted, traps must be checked early in the morning during the period when temperatures will have minimal effect on the trapped animals (no later than 3 hours after sunrise but as early as possible in high temperature conditions). If checking of traps cannot be completed within this timeframe, trap numbers must be reduced or the number of personnel increased before any further trapping occurs.

(c) Traps must either be closed on checking and re-opened late afternoon, or, if they need to remain open (i.e. targeting diurnal animals), the Animal Ethics application must provide information to show that leaving traps open during the day will not impact animal welfare. Traps remaining open during the day must be in a shaded position, and consideration should be given to more frequent checking throughout the day, particularly in hot weather or if there are non-target captures.

(d) An appropriate handling bag must be carried when approaching a trap to ensure rapid removal of the animal from the trap (see the Department SOP for *Animal Handling and Restraint using Soft Containment*).

(e) Take care when approaching the trap and minimise noise during your approach (reduce noise from walking, leaf crunch, vehicles, talking etc.). A second handling bag may be used to quickly cover the front of the trap (the exposed front of the trap) which may aid in calming agitated trapped animals, particularly woylies.

(f) Bait within each trap should be checked daily and replaced when necessary. Traps without bait reduce the validity of trapping results.

(g) The presence of ants in the trapping area can lead to detrimental impacts on captured animals. A small amount of surface insecticide (e.g. permethrin-based products such as Coopex) can be applied around and below traps to discourage ants. Liquid or spray

insecticides should never be used inside traps, but can be applied around the outside. Extreme care must be taken to ensure that no free standing liquid droplets remain when using liquid-based permethrin as absorption/ingestion can be lethal to frogs and reptiles. Always read the MSDS of chemicals before use. If ants become highly attracted to the trapping area, remove and relocate the traps to a more suitable position. One way to reduce risk of ant infestation is to remove bait each morning, when clearing traps and replace when resetting in afternoon.

**ANIMAL WELFARE:** If moderate to high numbers of ants are identified at a trap site, or if small numbers of ants cause welfare issues, then the trap must be closed or moved to another location.

(h) Trapping data should be recorded on an appropriate trapping datasheet and in a database.

## 5.4 Removing animals from traps

All animal handling must be done by (or under the guidance of) trained and competent personnel. Techniques for removing animals from traps vary depending on the species involved and the experience and skills of the animal handler. These notes are provided as a general guide only.

**ANIMAL WELFARE:** Capture myopathy is a condition associated with the capture and handling of many species of mammals and birds that results in degeneration of skeletal and/or cardiac muscle (Shepherd *et al.*, 1988). The condition can result in sudden death but death may also occur weeks after capture as a result of complications including abnormalities to posture and gait and increased susceptibility to predation (Abbot *et al.*, 2005). Signs and symptoms include a drooping head and neck, laboured breathing, tremors, lethargy and lack of coordination or paralysis.

Prevention of the condition through efforts to minimise stress to animals is better than treatment options. Records of animals suspected to be suffering from capture myopathy need to be provided to the Animal Ethics Committee for annual reporting requirements.

To ensure minimal stress to the animals, animals should only be handled for as long as required to identify them and to collect any necessary measurements (usually no more than five minutes). At a maximum they must be released (or reach alternate end point) within 24 hours of capture.

Ejection of pouch young is common in species of the Potoroidae and Peramelidae families. Persons that may encounter species of these families whilst trapping must be familiar with the Department SOP for *Care of Evicted Pouch Young*. Records need to be kept on orphans, their care and fate for annual reporting requirements for the Department's Animal Ethics Committee approved projects.

(a) Use handling bags appropriate for the species and length of containment as advised in Department SOP for *Animal Handling and Restraint using Soft Containment*.

**ANIMAL WELFARE:** All handling bags and equipment should be kept clean to minimise risk of disease, contamination, etc. Refer to the Department SOP for *Managing Disease Risk in Wildlife Management* for guidance.

(b) Remove animals from the trap as efficiently as possible.

- (c) Keep traps covered as much as possible during removal of the animal to minimise stress.
- (d) **Small Cage:** Animals should be encouraged to enter the handling bag by placing the bag over the end of the trap and manipulating the door to the open position. Lifting small cages with an animal inside should be avoided. Gentle encouragement via blowing on the animal (e.g. short, sharp breaths), using light and dark or positioning of the animal handler's body toward the rear of the trap can help.
- (e) Check for dependent young after adult is removed from trap (inside the back of the trap, under hessian and beside the trap).
- (f) Particular care should be taken for those species that may eject pouch young.
- (g) Venomous or dangerous animals such as snakes should be released with consideration given to the best possible escape route for both animal and personnel. The door can be propped open to allow the animal to leave when the animal is ready.
- (h) Captured animals must be released at point of capture (unless the purpose of the trapping is translocation, specimen collection is required or other approved reason). Animals should be released as soon as possible and at an appropriate time of day or night. Animals must be released, or reach an alternate endpoint approved by the Department's Animal Ethics Committee, within 24 hours of capture. Animals should be released into good shelter where necessary and caution taken to reduce exposure to risks such as predation.
- (i) Where practical, non-targets, particularly birds, should be assessed for injury.

## 5.5 Picking up traps

- (a) All traps must be counted out upon setting traps and counted in when picking up. Personnel undertaking the trapping should keep tallies of traps to ensure that all are collected and that there are no traps left behind. If traps are not being collected immediately after checking (i.e. traps are not being checked and picked up simultaneously), the traps must be closed on checking and remain closed until they are picked up. This is the responsibility of the person in charge at the survey location on the day. There is no excuse for leaving traps set in the field.
- (b) Ensure residual bait is removed from traps and flagging tape is removed from the area.

## 6 Trap Care and Maintenance

**ANIMAL WELFARE:** Traps and hessian covers must be cleaned and disinfected after each trapping session. Do not move dirty hessian covers and traps from one working site to another as it poses a disease risk for animal populations. To avoid possible transfer of pathogens use one batch of hessian covers and traps for each site or connected group of sites. Refer to the Department SOP for *Managing Disease Risk in Wildlife Management*.

- (a) Traps must be maintained in good working order.
- (b) In some instances, particularly traps that have held reptiles or brushtail possums, the trap will require faecal material to be removed within a trapping period. Particular attention should be paid to the release mechanism to ensure it is kept free of bait and scats. Instructions

on cleaning and disinfection of traps are available in the Department SOP for *Managing Disease Risk in Wildlife Management*.

(c) Hessian bags used as trap coverings should also be cleaned and disinfected after each trapping session following the instructions contained in the Department SOP for *Managing Disease Risk in Wildlife Management*.

(d) Do not carry the traps by any of the moving parts and do not put any excessive weight into traps that will be carried.

(e) Any damaged traps requiring attention need to be flagged and labelled in the field when a problem is identified so that it can be attended to and removed from use until repaired.

## 7 Competencies and Approvals

Department personnel, and other external parties covered by the Department's Animal Ethics Committee, undertaking monitoring projects involving cage traps require approval from the Committee and will need to satisfy the competency requirements detailed in Table 1. This is to ensure that personnel involved have the necessary knowledge and experience to minimise the potential impacts of cage trapping on the welfare of the animals. Other groups, organisations or individuals using this SOP to guide their fauna monitoring activities are encouraged to also meet these competency requirements as well as their basic animal welfare legislative obligations.

It should be noted that details such as intensity of the study being undertaken will determine the level of competency required and Table 1 provides advice for basic monitoring only.

*Table 1 Competency requirements for Animal Handlers of projects using cage traps to capture fauna*

Competency category	Competency requirement	Competency assessment
<b>Wildlife licences</b>	Licence to take fauna for scientific purposes (Reg 17) OR Licence to take fauna for educational or public purposes (Reg 15)	Provide licence number
<b>Formal training</b> <i>Note: Suitable levels of skills/experience can substitute for formal training requirements</i>	Department Fauna Management Course or equivalent training	Provide course year
<b>General skills/experience</b>	Relevant knowledge of species biology and ecology	Personnel should be able to correctly identify the likely species to be captured in cage traps for the site/s being studied. This knowledge may be gained through sufficient field experience and/or consultation of field guides and other literature.

Competency category	Competency requirement	Competency assessment
		Estimated total time in field: Min 1 year involved in similar projects.
<b>Fauna survey and capture skills/experience</b>	Experience in setting and use of live traps	Personnel should be confident identifying the best locations to set traps and how to set traps so that the mechanism works and animal welfare is considered at all times. This knowledge may be gained through sufficient field experience and/or consultation of literature. Estimated total time in field: Min 1 year involved in similar projects.
	Training and experience in trap hygiene, disease transmission	Personnel should be familiar with hygiene procedures. This knowledge may be gained through sufficient field experience and/or consultation of literature. Estimated total time in field: Min 1 year involved in similar projects.
<b>Animal handling and processing skills/experience</b>	Experience in handling terrestrial fauna	Personnel should be confident handling and restraining the range of species likely to be captured. This knowledge may be gained through sufficient field experience and/or consultation of literature. Estimated total time in field: Min 2 years involved in similar projects.

## 8 Occupational Health and Safety

Always carry a first aid kit in your vehicle and be aware of your own safety and the safety of others as well as the animals when handling.

A job safety analysis is recommended prior to undertaking any monitoring which involves hand capture. This safety analysis should include the following considerations.

### 8.1 Animal bites, stings and scratches

Care should be taken when handling animals to avoid bites, stings or scratches. All inflicted injuries (even superficial ones) should be appropriately treated as soon as possible to ameliorate possible allergic reaction, prevent infection and promote healing.

To improve safety, field personnel should be aware of the treatment for snakebite and carry appropriate pressure bandages. Personnel should also have up-to-date tetanus vaccinations. Department personnel must not capture bats unless fully vaccinated against Australian Bat Lyssavirus.

If Department personnel or volunteers are injured, please refer to the Department's Health and Safety Section's 'Report a Hazard, near-miss or incident' intranet page, which can be

found at [http://intranet/csd/People\\_Services/rm/Pages/ReportingHazards,Near-MissesandIncidents.aspxZoonoses](http://intranet/csd/People_Services/rm/Pages/ReportingHazards,Near-MissesandIncidents.aspxZoonoses).

## 8.2 Zoonoses

There are a number of diseases carried by animals, including ticks, that can be transmitted to humans (i.e. zoonoses such as Toxoplasmosis, Leptospirosis, Salmonella). All personnel must take precautions to minimise the risk of disease transmission to protect themselves, their families and wildlife populations.

Advice on minimising disease risk is contained in the Department SOP for *Managing Disease Risk in Wildlife Management*

## 8.3 Allergies

People with or that develop severe allergies associated with animals or animal materials should consult with their medical practitioner on appropriate precautions and actions for the handling of wildlife. People with nut allergies should demonstrate caution when handling, or in the vicinity of, universal bait.

## 8.4 Hygiene

Personnel should ensure that they maintain high standards of personal hygiene at all times, particularly in relation to handling bait/food materials. Inappropriate handling of food, especially meat products, can lead to illness and infection in personnel and in animals. Hands, equipment and anything that has come in contact with food items should be thoroughly washed and disinfected.

# 9 Further Reading

The following SOPs have been mentioned in this advice and it is recommended that they are consulted when proposing to use cage traps:

- Department SOP *Soft Cage Traps for Capture of Macropods*
- Department SOP *Animal Handling and Restraint using Soft Containment*
- Department SOP *Care of Evicted Pouch Young*
- Department SOP *Humane Killing of Animals under Field Conditions*
- Department SOP *First Aid for Animals*
- Department SOP *Managing Disease Risk in Wildlife Management*

For further advice refer also to:

Environmental Protection Authority and Department of Environment and Conservation (2010) *Technical Guide - Terrestrial Vertebrate Fauna Surveys for Environmental Impact Assessment* (Eds. B.M. Hyder, J. Dell and M.A Cowan). Perth, Western Australia: EPA and DEC.

# 10 References

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## 11 Glossary of Terms

**Animal handler:** A person listed on an application to the Department's Animal Ethics Committee who will be responsible for handling animals during the project.

**Cage trap:** A trap for the live capture of animals constructed of wire mesh. Cage traps operate by the animal treading on a weight-sensitive trigger plate which causes the door to close and lock.

# Appendix I: Universal Bait Recipe

## Equipment

- Mixing bowl or bucket
- Mixing spoon (optional: can just use your hands)
- Container with lid to store bait
- Disposable gloves

## Ingredients

- 500g Quick cooking oats
- 2 kg (5-6 375g tubs) Smooth peanut butter
- *Optional:* Between 110g (1 tin) and 636g (6-8 tins) Sardines (preferably in oil, or springwater)
- *Optional:* Cooking oil, preferably peanut oil

*Note: Avoid using ingredients that contain additives, preservatives or artificial colours and flavours.*

Serves: makes enough bait for approximately 100 cage traps for 4 trap nights.

## Methodology

1. Ensure staff mixing bait are not allergic to peanuts.
2. Place oats and sardines into clean mixing bowl or bucket and mix so that the sardines are well distributed through the oats.
3. Mix in peanut butter until the oats and sardines are well distributed and the mixture is not too dry or too sticky. Form a ball that is sticky and cohesive. Keep in mind that the mixture will become drier over time as the oats absorb the oil from the peanut butter.
4. Store bait in a sealed container.
5. Clean bait mixing equipment.
6. Add extra peanut butter if mixture becomes too dry. Water or cooking oil can be used if extra peanut butter is not available.

*Optional:* Bait can be pre-rolled.

Roll bait into balls ready for placing in traps (approx. 20c coin size for cage traps and 10c coin size for box traps). The bait balls can be counted to match the number of traps being set. This will ensure that you have enough bait for all traps being set and will also act as an additional check to ensure all traps have been set and baited.

## Animal Welfare

To reduce the risk of impact of the use of universal bait on wildlife ensure that bait is stored for no longer than the specified period of 5 days fresh, or 3 months frozen, to avoid the risk of the components spoiling and unsuitable for consumption. Where possible do not leave bait in open sun. Any old bait should be disposed of after trapping and not frozen for later reuse. Do not use old bait or bait containers that have mouldy bait in them.

Potential animal welfare impacts of mixing universal bait include:

- Food poisoning
- Changing behaviour by providing a food source

## References

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- Wayne, A.F., Rooney, J., Morris, K.D., Johnson, B., 2008. Improved bait and trapping techniques for chuditch (*Dasyurus geoffroii*): overcoming reduced trap availability due to increased densities of other native fauna. *Conservation Science Western Australia* 7(1):49-56.
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